

RIPA Lysis Buffer (Strong)

Catalog# BWR1004

Size: 100 ml

Lot # Check on the product label

Introduction

1. This RIPA Lysis Buffer is a kind of traditional cell and tissue lysis buffer. The end protein / sample lysed by this buffer can be used for Western Blot and Immunoprecipitation.
2. RIPA (Radio Immunoprecipitation Assay) Lysis Buffer has different formulas, and according to its lysis strength, it can be roughly classified into three categories: strong, medium and weak.
3. RIPA Lysis Buffer (Strong) contains: 50mM Tris (pH 7.4), 150mM NaCl, 1% Triton X-100, 1% sodium deoxycholate, 0.1% SDS, and different kinds of inhibitors (sodium orthovanadate, sodium fluoride, EDTA and leupeptin), which can effectively inhibit the protein degradation.
4. The concentration of the protein which lysed by this buffer can be determined by our BCA Protein Assay Kits (Catalog # BWR1023, BWR1024, BWR1025, BWR1026). For the high concentration of detergent, it is not recommended to use Bradford Protein Assay Kit.

Kit Components

Components	Size	Storage Instruction
RIPA Lysis Buffer (Strong)	100 ml	Store at 4°C for one year

Protocol

● Cultured Cells Sample

1. Pipette proper volume of RIPA Lysis Buffer and mix thoroughly. 2-3 min advanced before use, add PMSF buffer to make PMSF final concentration to 1mM.

2. For **adherent cells**: wash sample with PBS, normal saline or serum-free culture medium to remove culture solution. Add proper volume of RIPA Lysis Buffer, then stroke with pipette until the buffer immerse cells completely. Shake slightly for 5-10 min. After lysis, centrifuge at 10,000-14,000 g for 10 min, then collect the supernatants and move on to the next step.

>> Instruction for RIPA usage: for different sizes of cell culture plates <<

Sizes of plate / surface area	Volume of Buffer
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Chongqing Biospes Co., Ltd Tel: +86-23-67567091 Fax: +86-23-67745923

7F, Bldg B, High-tech Venture Park, # 107 Erlang Chuangye Rd, Jiulongpo District, Chongqing, 400039, China

www.biospes.com

100 mm	500-1,000 μ l
60 mm	250-500 μ l
6-well plate	200-400 μ l per well
24-well plate	100-200 μ l per well
96-well plate	50-100 μ l per well

3. For **suspending cells**: Centrifuge to collect cells, then, wash sample with PBS, normal saline or serum-free culture medium. Add proper volume of RIPA Lysis Buffer, then stroke with pipette until cells dispersed. Vertex for 5-10 min to lyse cells completely (There should be no obvious cell precipitates if cells lysed completely. For large volume of cells, aliquot and lyse then). After lysis, centrifuge at 10,000-14,000 g for 10 min, collect the supernatants and move on to the next step.

● Tissues Sample

1. Put the tissue sample into precooling normal saline quickly, remove blood by rinsing it several times. Weight sample and cut it into small slices, then put them into the tissue homogenizer.

2. Pipette proper volume of RIPA Lysis Buffer and mix thoroughly. 2-3 min advanced before use, add PMSF buffer to make its final concentration to 1mM.

3. Tissue net weight (g) to RIPA Lysis Buffer (ml) ratio = 1:10 (i.e. add 10ml of RIPA Lysis Buffer to 1g tissues) and homogenate (If lyses incompletely, can add more volume of RIPA Lysis Buffer; if high concentration protein samples are required, can reduce the volume of lysis buffer).

4. Homogenize with a glass homogenizer until samples were lysed completely.

5. Centrifuge at 10,000-14,000 g for 3-5 min, collect the supernatant and move on to next step.

Note: It is normal to see a small group of transparent jelly in the end product of RIPA Lysis Buffer. This transparent jelly is a complex containing genomic DNA. If not detect the genomic DNA binding proteins, users can centrifuge directly and collect the supernatant to do next steps of the experiment; Or, scatter the jelly by ultrasonic wave, then, centrifuge and collect the supernatant to do next steps of the experiment. For detecting the normal transcription factors (e.g. NF-kappa B, P53, etc.), ultrasonic wave processing is not necessary.

Notes

1. All steps of protein extraction should be operated on ice or at 4 $^{\circ}$ C . It is recommended to aliquot the sample into sub-packages at proper volume, then freeze-drying or store at -20 $^{\circ}$ C in liquid form. Avoid freeze thawing repeatedly.

2. PMSF (phenylmethylsulfonyl fluoride) buffer is not included in this product, we provide it separately (Catalog # BWR1014). Few minutes (2-3 min) advanced before use, add PMSF buffer to make its final concentration to 1mM.

3. Please wear the lab coat and disposable gloves to operate.

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